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(54) Continuous release pharmaceutical compositions

Pharmazeutische Zusammensetzungen für die kontinuierliche Freigabe des Wirkstoffes

Compositions pharmaceutiques pour la libération continue de la substance active

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Description

[0001] This invention relates to solid pharmaceutical compositions of the pharmacologically-active acid-stable polypeptide ICI 188,630 which provide continuous release of the polypeptide over an extended period when the composition is placed in an aqueous, physiological-type environment

[0002] It has long been appreciated that the continuous release of certain drugs over an extended period following a single administration could have significant practical advantages in clinical practice, and compositions have already been developed to provide extended release of a number of clinically useful drugs, after oral dosing (see, for example, Remington's Pharmaceutical Sciences, published by Mack Publishing Company, Easton, Pennsylvania, U.S.A., 15th Edition, 1975, pages 1618—1631), after parenteral administration (*ibidem*, pages 1631—1643), and after topical administration (see, for example, United Kingdom Patent Number 1,351,409). A suitable method of parenteral administration is the sub-dermal injection or implantation of a solid body, for example a pellet or a film, containing the drug, and a variety of such implantable devices has been described. In particular, it is known that, for many drugs, suitable implantable devices for providing extended drug release may be obtained by encapsulating the drug in a biodegradable polymer, or by dispersing the drug in a matrix of such a polymer, so that the drug is released as the degradation of the polymer matrix proceeds.

[0003] Suitable biodegradable polymers for use in sustained release formulations are well known, and include polyesters, which gradually become degraded by hydrolysis when placed in an aqueous, physiological-type environment. Particular polyesters which have been used are those derived from hydroxycarboxylic acids, and much prior art has been directed to polymers derived from α -hydroxycarboxylic acids, especially lactic acid in both its racemic and optically active forms, and glycolic acid, and copolymers thereof—see, for example, United States Patents Numbers 3,773,919 and 3,887,699; Jackanicz *et al.*, Contraception, 1973, 8, 227—234; Anderson *et al.*, *ibidem*, 1976, 11, 375—384; Wise *et al.*, Ufe Sciences, 1976, 19, 867—874; Woodland *et al.*, Journal of Medicinal Chemistry, 1973, 16, 897—901; Yolles *et al.*, Bulletin of the Parenteral Drug Association, 1976, 30, 306—312; Wise *et al.*, Journal of Pharmacy and Pharmacology, 1978, 30, 686—689 and 1979, 31, 201—204.

[0004] In this specification, the term "polylactide" is used to include copolymers of lactic acid and glycolic acid as defined in claim 1 and mixtures of such copolymers, the lactic acid being either in racemic or in optically active form. Also, the term "acid-stable" is to be understood as meaning that the polypeptide is not significantly hydrolysed under the conditions encountered within the claimed formulations during the period of use envisaged, that is, at pH 2 at mammalian body temperature, say up to 40°C, for up to six months.

[0005] United Kingdom Patent Specification Number 1,325,209 (equivalent to United States Patent Specification Number 3,773,919) and United States Patent Specification Number 3,887,699 is the only prior art known to us which makes any reference to extended or sustained release of polypeptides, the latter mentioning insulin only, but it contains no specific example of any such formulation, and the reference to polypeptides is apparently entirely speculative, appearing, as it does, only in an extensive listing of very many different classes of drugs which can allegedly be incorporated into formulations of the kind described therein. In fact, essentially all of the other drug types referred to in that specification, apart from polypeptides, are relatively hydrophobic in character and of relatively low molecular weight, and the disclosure of that specification displays no recognition of the difficulties which we have encountered when seeking to obtain satisfactory sustained release formulations of polypeptides, many of which are relatively hydrophilic, and of relatively high molecular weight

[0006] It is to be appreciated that "sustained" or "extended" release of a drug may be either continuous or discontinuous. We have now discovered, in fact, that in many cases when the teaching of the prior art, and in particular the teaching of United Kingdom Specification No. 1,325,209, is applied to the manufacture of a formulation of an acid-stable polypeptide, the release of the polypeptide from the formulation, although occurring over an extended period of time, may also be discontinuous. For example, the release of a polypeptide from a polylactide polymer as described in the said Specification is often preceded by a significant induction period, during which no polypeptide is released, or is biphasic, and comprises an initial period during which some polypeptide is released, a second period during which little or no polypeptide is released, and a third period during which most of the remainder of the polypeptide is released. By contrast, it is an object of the present invention to provide compositions of the acid-stable polypeptide from which, apart possibly from a relatively short initial induction period, the polypeptide is released continuously, with no periods during which little or no polypeptide is released. The words "continuous release" are used in this specification solely to describe a release profile which is essentially monophasic, although it may have a point of inflection, but certainly has no "plateau" phase.

[0007] In Journal of Bioengineering, 1, 25—29 (1976) (Chemical Abstracts, 88, 7904/n, 1978), two formulations of insulin are disclosed which show an essentially monophasic release profile, but substantially all the insulin is released within about 48 hours.

[0008] Thus, according to the present invention, there is provided a solid pharmaceutical composition as defined in claim 1.

[0009] By "an aqueous physiological-type environment" we mean the body, particularly the musculature or the circulatory system, of a warm-blooded animal, although in laboratory investigations such an environment may be mimicked by aqueous liquids, optionally buffered to a physiological pH, at a temperature of between 35 and 40°C.

[0010] The continuous release composition of the invention may be placed in the body of an animal which it is desired to treat with the polypeptide by, for example, intramuscular or subcutaneous injection, or by sub-dermal surgical implantation, in conventional clinical or veterinary manner.

[0011] We have discovered that a pharmaceutical composition according to the invention may be designed to provide continuous release of the polypeptide by the appropriate choice or control of various parameters, for example by varying the polylactide composition, particularly the proportion of lactic acid to glycolic acid in copolymers; by controlling the molecular weight of the polylactide, both the weight-average molecular weight, and the molecular weight range of polydispersity, measured by the ratio of the weight-average molecular weight, (M_w), to the number-average molecular weight (M_n), i.e.

$$\frac{M_w}{M_n}$$

by choice of the proportion of polypeptide to polylactide; or by choice of the geometry of a solid formulation for implantation, or of the particle size in a formulation for injection. The release characteristics of such compositions are also controlled to some extent by the nature of the polypeptide itself. In particular, there is less freedom of choice in defining the parameters mentioned above when designing a composition of the invention for a polypeptide of high molecular weight, (say, greater than 6000), than there is when designing a composition for a polypeptide of lower molecular weight, (say, less than 6000).

[0012] We have further discovered that the release of polypeptide from a composition comprising polylactide and polypeptide proceeds by two distinct and independent mechanisms, namely first a diffusion-dependent release of polypeptide out of the polylactide-polypeptide matrix comprising leaching from the surface, and, for low molecular weight polypeptides, some partition-dependent diffusion of polypeptide per se; and subsequently, as the polylactide becomes degraded, diffusion of aqueous polypeptide solution out of the composition through aqueous channels.

[0013] In general, the compatibility of polypeptides in polylactide polymers is limited, except in the case of low molecular weight (up to, say, 6000 molecular weight) polypeptides which are capable of having some specific interaction with the polylactide, for example a low molecular weight polypeptide which is basic, and which therefore interacts with the terminal carboxylic acid groups in the polylactide. Because of this limited compatibility of polypeptide in polylactide, a polypeptide/polylactide formulation, when placed in an aqueous environment, releases very little polypeptide by diffusion through the polymer matrix. While this is broadly true for all combinations of polypeptide and polylactide, matrix diffusion is at a minimum for high molecular weight polypeptides in high molecular weight polylactides. Even when some matrix diffusion resulting in polypeptide release occurs initially on placing the composition in an aqueous environment, by release, from or very near the surface, this soon ceases because the diffusion of polypeptide into polylactide is insufficient to result in continuous transport of polypeptide from inside the composition to its surface.

[0014] When a polypeptide/polylactide composition is placed in an aqueous environment, water diffuses into the matrix, and is partitioned between polypeptide and polylactide to form domains of aqueous polypeptide solution. This aqueous polypeptide, obtained when absorbed water is partitioned between polypeptide and polylactide, is incompatible with, and insoluble in, polylactides, particularly those of high molecular weight, so that absorption of water by the composition still further reduces initially any likelihood of matrix diffusion of polypeptide out of the composition. If the aqueous domains of polypeptide so formed are discrete and isolated, then the compositions are incapable of releasing polypeptide. However, the possibility of the aqueous polypeptide domains having some continuity increases with increasing concentration of polypeptide in the composition, and with increasing absorption of water, and when the continuity of aqueous polypeptide domains reaches a sufficient level to communicate with the exterior surface of the composition, polypeptide starts to be released from the formulation by diffusion, not through the polylactide matrix, but through aqueous polypeptide channels. Even when some aqueous polypeptide domains near the surface have extended so as to reach the exterior of the composition, that aqueous polypeptide which exists in still isolated domains is not released, and is only released when a secondary hydrophilic path for diffusion becomes available. For high molecular weight polylactides, this secondary hydrophilic diffusion path arises when the polylactide has undergone sufficient degradation for the rate of absorption of water to increase significantly. When this occurs, aqueous pores or channels are generated in the polylactide matrix, which allow a continuous and significant release of polypeptide, as aqueous solution, from previously discrete and isolated domains.

[0015] As indicated above, we have discovered that, when sustained release compositions of the prior art, and particularly those described in United Kingdom Patent Specification Number 1,325,209, are used for the release of polypeptides, the initial matrix diffusion phase of polypeptide release, and the secondary release of polypeptide con-

sequent upon degradation of the polylactide, are separated in time with the result that the release of polypeptide is not continuous, but is biphasic and discontinuous, comprising a small first release of polypeptide, a dead phase during which essentially no polypeptide is released, and subsequent second release phase, during which substantially all of the remaining polypeptide is released. We have now discovered that, by appropriate choice of the parameters of the composition, the matrix diffusion phase of release, and subsequent degradation induced phase of release, can be made to overlap in time.

[0016] The two phases of release can be made to overlap by either extending the initial diffusion phase, or making the degradation-induced phase commence earlier, or both.

[0017] The initial matrix release phase is difficult to extend, but is sensitive to the concentration of the polypeptide in the matrix, and to a limited extent, to the nature of the polypeptide, especially its hydrophilicity.

[0018] The degradation-induced release phase can be made to start sooner by appropriate choice of polylactide composition (more glycolide-rich polymer molecules, which degrade more quickly than lactide-rich molecules), M_w (molecules of low molecular weight degrade more quickly to a level at which aqueous channels appear in the matrix), and polypeptide concentration (a higher polypeptide concentration allows more rapid absorption of water, and consequently more rapid generation of continuous aqueous channels which facilitate polypeptide release).

[0019] However, as well as requiring the degradation induced release phase to start earlier, it is also necessary to control the rate of polypeptide release during this phase, and to ensure that the total duration of this phase is sufficient for its intended clinical or veterinary purpose. One method of extending the duration of the degradation-induced release phase, is to use polylactides which contain lactide-rich molecules, which degrade more slowly than glycolide-rich molecules, or alternatively, polylactides containing molecules of high molecular weight, which take longer to degrade to a level at which aqueous channels are formed, can be used.

[0020] It is apparent, therefore, that since glycolide-rich polymer molecules, and/or molecules of low molecular weight, are preferred if the degradation phase is to start quickly, and lactide-rich molecules, and/or molecules of high molecular weight are preferred if the degradation-induced release phase is to last for a sufficient period of time, preferred polylactides are those with a high degree of heterogeneity, in respect of glycolide-rich and lactide-rich molecules, or of high polydispersity.

[0021] Alternatively, the same characteristics can be obtained by blending two or more different polylactides, which differ in lactide/glycolide content, and/or in M_w . In addition, the blending of a minor proportion of polylactide of high M_w with a polylactide of low M_w , confers desirable physical properties upon compositions according to this invention produced therefrom, making them easier to fabricate and process.

[0022] We have further discovered that the profile of polypeptide release from both prior art polylactides and novel polylactides is almost exactly paralleled by the profile of water absorption. That is to say, when polypeptide release is discontinuous, water absorption is also discontinuous in essentially the same manner, and conversely, when polypeptide release is continuous, so is water absorption. Furthermore, variation of the parameters referred to above for controlling the polypeptide release characteristics of the composition have been found to affect water absorption by the composition in an exactly parallel manner.

[0023] The effect of the various parameters, referred to above, on the polypeptide release and/or water absorption characteristics of compositions of the invention is illustrated by the following experiments, which do not necessarily illustrate compositions of the invention.

A. Molecular weight of the polylactide component

A.1. Low molecular weight polypeptide.

A.1.1. Formulations were manufactured comprising 20% w/w of the gastric peptide fragment tetragastrin hydrochloride, Trp-Met-Asp-Phe-NH₂·HCl, molecular weight=633, in a polylactide comprising equimolar proportions of D,L-lactide and glycolide units, in the form of a film 0.2 mm thick. The films were placed individually in water at 37°C, which was changed daily, and the ultraviolet absorption at 277 nm was measured to assay the tetragastrin released by the formulation that day.

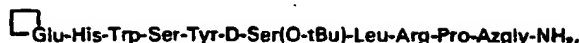
With a prior art type of polylactide of M_w approximately 240,000 (intrinsic viscosity=1.36), there was an initial release of tetragastrin, then a "dead period" from about day 5 to day 21 during which little was released, followed by the main release from day 24 onward.

With a polylactide of M_w approximately 15,000 (intrinsic viscosity=0.25), the release pattern was similar, but the dead period lasted only from day 4 or 5 to day 8 or 9.

With a polylactide of low M_w (inherent viscosity of a 1 g/100 ml solution in chloroform=0.11), there was no dead period, and tetragastrin was released continuously from time zero (T_0).

A.1.2. Similar formulations were manufactured using 10% by weight of the synthetic luliberin analogue

ICI.118,630,



molecular weight=1269, in place of tetragastrin.

With a prior art polylactide of $M_w \sim 240,000$ (intrinsic viscosity=1.36), the release of the polypeptide was biphasic, with a dead period of about 15 days.

With a polylactide of $M_w \sim 15,000$ (intrinsic viscosity=0.25), there was a short induction period, followed by continuous release.

With a polylactide of low M_w (inherent viscosity of a 1 g/100 ml solution in chloroform=0.11), there was continuous release from T_0 .

A.2. Medium molecular weight polypeptide.

Formulations were manufactured comprising 0.1% by weight of mouse epidermal growth factor (EGF), molecular weight=6041, in polylactides comprising equimolar proportions of D,L-lactic acid and glycolic acid units, and of different M_w , and placed in pH 7.4 buffer. Release of EGF was monitored by radio-immunoassay.

With a prior art polylactide of $M_w \sim 200,000$ (intrinsic viscosity=1.08), there was no initial release, and no significant release of polypeptide until between 13 and 20 days after T_0 , after which release was continuous.

With a polylactide of $M_w \sim 80,000$ (intrinsic viscosity=0.58), there was also no initial release, and significant release did not occur until between 6 and 10 days after T_0 , after which release was continuous.

With a polylactide of low M_w (inherent viscosity of a 1 g/100 ml solution in chloroform=0.11), there was continuous release from T_0 .

A.3. High molecular weight polypeptide

A formulation was manufactured comprising 20% w/w of bovine prolactin, molecular weight $\sim 22,000$, in a polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units, and of low M_w (inherent viscosity of a 1 g/100 ml solution in chloroform=0.11). As expected from previous experiments A.1.1., A.1.2. and A.2., this formulation also released polypeptide continuously from T_0 when tested *in vivo* in rats, and circulating bovine prolactin assayed by radio-immunoassay.

Experiments A.1. to A.3. thus demonstrate that reducing the molecular weight, M_w , or viscosity of the polylactide used in the manufacture of a composition reduces the biphasicity of polypeptide release, for polypeptides of low or medium molecular weight, or the initial delay in release of polypeptides of medium or high molecular weight, and achieves a continuous release of the polypeptide from T_0 .

B. Ratio of lactide/glycolide in the polylactide

Compositions were manufactured in the form of implants containing 100 μ g (3% w/w) of the luliberin analogue ICI. 118,630 in polylactides of $M_w \sim 300,000$ but of different lactide/glycolide ratios. All of these formulations when tested *in vivo* in adult female rats exhibiting normal oestrous behaviour, gave biphasic release of the polypeptide, comprising release for about 6 days post-treatment, followed by a dead period, during which there was no significant polypeptide release. The length of this dead period decreased with decreasing lactide/glycolide ratio (L/G), as follows:—

L/G	Dead period (days)
100/0	no release
75/25	51
67/33	34
50/50	15

This experiment thus indicates that a composition exhibiting a biphasic release of polypeptide can be improved in the direction of achieving continuous release by increasing the proportion of glycolide to lactide in the polylactide used, up to about 50% glycolide.

C. Ratio of polypeptide to polylactide

Compositions in the form of implants were manufactured using different concentrations of the synthetic luliberin analogue, ICI.118,630, in a prior art 50/50 lactide/glycolide polylactide of $M_w \sim 200,000$, and tested *in vivo* in rats

as described above. At 5% and 10% w/w incorporation, the release of polypeptide was biphasic, but at 15% and 20% incorporation, the biphasicity disappeared.

This experiment thus demonstrates that a polylactide of high molecular weight, which gives biphasic release of polypeptide when the polypeptide is incorporated only at low levels, can be used to make satisfactory continuous release formulations if the proportion of polypeptide is increased sufficiently.

D. Molecular weight distribution

A solution of a polymer blend of wide molecular weight distribution (polydispersity) was obtained by mixing solutions of a 50/50 D,L-lactide/glycolide polylactide of low M_w (reduced specific viscosity of a 1 g/100 ml solution chloroform=0.115), (3 parts by weight) and a 50/50 D,L-lactide/glycolide polylactide of $M_w=200,000$, (intrinsic viscosity=1.08), (1 part by weight). Tetragastrin (1 part by weight) was added, and the mixture was cast to give a polylactide/tetragastrin composition containing 20% by weight of tetragastrin, and this was then moulded to give a slab 0.02 cm thick. The slab was placed in water at 37°C and the release of tetragastrin was found to be continuous from T_0 , and to continue for at least 44 days.

Polylactides of wide molecular weight distribution may be obtained either by mixing preformed polymers of different molecular weights, or by appropriate control of the polymerisation process in generally known manner, and such polylactides confer important advantages, for example the lower molecular weight polylactide species allow an essentially immediate release of some polypeptide, while the higher molecular weight polylactide species both extend the release period and slow down the overall rate of release of the polypeptide. In addition, the blending of low and high M_w fractions modifies the water absorption characteristics of the polylactide in a parallel manner.

E. Thickness of the implant

E.1. A solution of 10% by weight of tetragastrin in a polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units, and of $M_w \sim 15000$ was cast into films 0.02, 0.06 and 0.12 cm thick. All three films showed continuous release of tetragastrin from T_0 , and at 28 days, the three films had released respectively 85, 75 and 66% of their tetragastrin content.

E.2. The uptake of tritiated water from pH 7.4 buffer by slabs of polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units of $M_w \sim 200,000$, and intrinsic viscosity=1.08 and of thickness 0.02, 0.06, 0.12 and 0.20 cm was measured by removing such slabs from the buffer solution successively after varying times of immersion, and measuring the tritium content by scintillation counting. After 5 weeks, the different thickness slabs had absorbed respectively 44, 20, 15 and 11% by weight of water.

These experiments show how the thickness of the implant can be used to control the absorption of water by, and thus the rate of release of polypeptide from, a composition of the invention, thicker implants releasing the polypeptide more slowly than thinner ones.

[0024] As indicated above the composition of the invention is a solid composition for sub-dermal injection or implantation or as a liquid formulation for intramuscular or subcutaneous injection.

[0025] Suitable solid compositions for sub-dermal injection or implantation are, for example, rods, spheres, films or pellets, and cylindrical rods which can be injected through a hypodermic needle or trochar are preferred. Such compositions are manufactured by conventional techniques which are well known in the pharmaceutical art.

[0026] Preferred solid compositions of the invention are as shown in Table 1. Each entry in Table 1 thus defines a further feature of this invention.

TABLE 1

No.	Polypeptide	Polylactide inherent viscosity*	$\frac{\text{Glycolide}}{\text{Lactide}}$	% Polypeptide
1	ICI.118630	0.2—0.5	0.2—3	5—50
2	ICI.118630	<0.2	0—3	0.1—50

* dl/g (1 g per 100 ml in chloroform or dioxan).

[0027] From the foregoing, it is clear that it is desirable to manufacture polylactides of a range of M_w , particularly of low to medium M_w in the range up to 60,000, and of high polydispersity

$$\frac{M_w}{M_n}$$

5 these being particularly valuable in the compositions of this invention. The prior art relating to polylactides in general, and to copolymers containing lactic acid and glycolic acid units in particular, is largely silent as to the manufacture of such copolymers of low molecular weight, and as to methods of achieving high polydispersity in such copolymers. Indeed, it is inherent in the prior art disclosure of polylactides that they are generally of M_w greater than about 30,000—60,000 (an inherent viscosity of greater than 0.5) and of low polydispersity, due to their manufacture under
10 anhydrous conditions and without any added chainstoppers.

[0028] Certain polylactides within this range have, however, been disclosed. For example, United States Patent Number 4,011,312 discloses copolymers having molecular weights in the range 1200—1800, and Polymer, 20, 1459—1464 (1979), United States Patent Number 3,773,919, United Kingdom Patent Number 1,351,409 and Chemical Abstrats., 91, 9442 g (1979) discloses polymers having a molecular weight upward of about 35,000, corresponding to
15 an inherent viscosity of greater than about 0.5.

[0029] We have realised that, because of the different reactivities under polymerisation conditions of the cyclic dimers of lactic acid and glycolic acid, copolymers of high heterogeneity in respect of polymer species may be obtained by ring opening polymerisation of a mixture of the two cyclic dimers in the presence of chain-stopping agents, to give polylactides having an inherent viscosity of less than 0.5. The cyclic dimer of glycolic acid is the more reactive under
20 polymerisation conditions, and thus the first copolymer molecules formed in the polymerisation are glycolic acid-rich. Consequentially, the later copolymer molecules formed are necessarily lactic acid-rich, thus producing a copolymer of lactic acid and glycolic acid of the desired high heterogeneity.

[0030] In addition, we control the polymerisation to produce copolymers in the desired low M_w range by carrying out the ring opening copolymerisation of the mixed cyclic dimers in the presence of water, of lactic acid containing water, or of some other known chain growth regulator, in accordance with the general knowledge in the polymer art.
25

[0031] Suitable polymerisation catalysts are zinc oxide, zinc carbonate, basic zinc carbonate, diethylzinc, organotin compounds, for example stannous octanoate, tributylaluminium, titanium, magnesium or barium compounds, or lithium, and of these stannous octanoate is preferred.

[0032] The copolymerisation of the mixed cyclic dimers is otherwise carried out in conventional manner, known in
30 the polymer art, as regards time and temperature.

[0033] According to a preferred feature of the invention the polylactide is a heterogeneous polymer containing from 25 to 100% molar of lactic acid units and from 0 to 75% molar of glycolic acid units, and which has an inherent viscosity (1 g/100 ml solution in chloroform or dioxan) equal to or greater than 0.09 and has an intrinsic viscosity equal to or less than 0.33. By a "heterogeneous polymer" we mean polymers with a high degree of heterogeneity, in respect of
35 glycolide-rich and lactide-rich molecules, or of high polydispersity, or blends of two or more different polylactides which differ in lactide/glycolide content and/or M_w , as hereinbefore described.

[0034] Whether or not a particular copolymer is heterogeneous or not, in this sense, may be readily determined from inspection of the 25 MHz ^{13}C nuclear magnetic resonance spectrum of the copolymer in, for example, deuterated dimethyl sulphoxide. In a homogeneous copolymer, such as is obtained in the prior art copolymerisation of lactic acid and glycolic acid monomers, the resonance of the glycolic acid unit carbonyl carbon, at $\delta = 166.0—166.2$ approximately, appears as two doublets, as a consequence of the four different, approximately equally probable molecular environments in which this carbon atom can exist, namely G^*G , L^*G , G^*L and L^*L (G=a glycolic acid unit, L=a lactic acid unit, and the asterisk indicates the glycolic acid unit under consideration). In a heterogeneous copolymer, on the other hand, such as is used in the present invention, the sequence LGL is unlikely to occur, so that one of the doublet signals
40 in the spectrum of the homogeneous copolymer appears as a singlet. In fact, we find that in the spectrum of heterogeneous copolymers, this glycolic acid unit carbonyl carbon signal often appears as two singlets. Thus, a "heterogeneous copolymer" as herein defined is a copolymer for which the glycolic acid carbonyl carbon signal in the ^{13}C n.m.r. appears as other than a pair of doublets.

[0035] The heterogeneity or homogeneity of lactic acid/glycolic acid copolymers can also be demonstrated by an examination of their degradation. Thus, when a copolymer is placed in pH 7.4 buffer at 37°C, removed periodically, dried and sampled, and the ratio of lactic acid to glycolic acid units in the samples is determined by n.m.r., for a heterogeneous copolymer, the ratio L/G increases with time, as the glycolic acid sequences hydrolyse preferentially. For a homogeneous copolymer, on the other hand, the ratio of L/G remains essentially constant as degradation progresses.
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[0036] The lactic acid content of the copolymer is preferably in the racemic (D,L) form, or in the optically active L form.

[0037] According to the invention the polylactide is made by a process, which comprises the ring opening copolymerisation of the cyclic dimer of lactic acid and in the presence of the cyclic dimer of glycolic acid, at elevated temperature in the presence of one of the above-mentioned polymerisation catalyst and in the presence of a chain stopping agent.
55

[0038] A suitable chain stopping agent is, for example, water, lactic acid, glycolic acid or other hydroxy acids, alcohols

or carboxylic acids generally.

[0039] The invention is illustrated but not limited by the following Preparations and Examples:—

Preparation 1

[0040] Zinc oxide (16 g) was added to D,L-lactic acid (800 g) in a 2 l3-necked round bottom flask-equipped with a stirrer, a thermometer, and a distillation head connected to a water condenser. The mixture was stirred and heated to about 135°C, at which temperature water started to distil over. Heating was continued for 8 h, during which time the temperature rose to about 190°C. When distillation of water ceased, the pressure was reduced, and distillation was continued until solid began to collect in the condenser. At this stage the water condenser was replaced by an air condenser, and the residue was cooled and then distilled under high vacuum [2.66—10.64 mbar (2—8 mm of mercury)], the fraction (about 300 g) distilling between 130 and 160°C being collected, this being D,L-lactide (3,6 - dimethyl -1,4 - dioxan - 2,5 - dione), the cyclic dimer of D,L-lactic acid.

[0041] The crude D,L-lactide was crystallised from ethyl acetate (approximately 600 ml) three times, and the recrystallised product was finally dried at 45°C under reduced pressure [2.66 mbar (2 mm of mercury)] for 24—48 hours, after which it had m.p. 124—125°C.

Preparation 2

[0042] Glycolide (1,4-dioxan-2,5-dione), the cyclic dimer of glycolic acid, was prepared by the method described in Preparative Methods in Polymer Chemistry by W. R. Sorenson and T. W. Campbell, second edition, published by Interscience (1968), page 363. The crude glycolide was purified by three successive crystallisations from dry ethyl acetate, then dried at 45°C under reduced pressure [2.66—10.64 mbar (2—8 mm of mercury)] for 24—48 h, m.p. 82—84°C.

Examples 1 to 13

[0043] Polymers of D,L-lactide and glycolide were prepared as follows:—

[0044] Pure dry D,L-lactide (Preparation 1), pure dry glycolide (Preparation 2) totalling 42 g, commercial D,L-lactic acid containing about 12% by weight of water, and 1 ml of an 8% by weight solution of stannous octanoate in hexane, were placed in a pre-dried glass tube. The hexane was evaporated under reduced pressure, and the tube was heated at 160°C for 6 hours with constant agitation if possible. The tube was cooled in powdered solid carbon dioxide, and the polylactide was removed, broken into small pieces and dissolved in chloroform (400ml). The chloroform solution was filtered, and the filtrate was poured into methanol (2 l) to precipitate the polylactide, which was filtered off and dried under vacuum at 40°C for 24 hours, then at 80°C for 24 hours. All the polylactides so produced were soluble in chloroform and dioxan, and polylactides 1 to 9 in the following Table were soluble in benzene, but polylactides 10 to 13 were insoluble in benzene.

[0045] The following particular polylactides were prepared by this method:—

Ex.	D,L lactide (L) (g)	Glycolide (G) (g)	L/G Molar proportion	D,L-lactic acid	Intrinsic viscosity (dl/g)	M _w (approx.)
1**	42.0	0	100/0	0	1.385	440,000
2**	33.5	9.0	75/25	0	1.084	400,000
3	32.4	8.7	75/25	920 µl	0.108*	low
4**	30.0	12.1	67/33	0	0.97	370,000
5**	30.0	12.1	67/33	0	0.94	214,000
6**	30.0	12.1	67/33	30 µl	0.67	107,000
7**	30.0	12.1	67/33	60 µl	0.51	63,000
8	30.0	12.1	67/33	120 µl	0.37	33,000
9	30.0	12.1	67/33	920 µl	0.121*	low
10**	23.0	18.5	50/50	0	1.045	300,000
11	23.0	18.5	50/50	400 µl	0.25	15,200

M_w are relative to polystyrene standard

* are reduced specific viscosities of a 1 g/100 ml solution in chloroform.

** are outside the scope of the invention

(continued)

Ex.	D,L lactide (L) (g)	Glycolide (G) (g)	L/G Molar proportion	D,L-lactic acid	Intrinsic viscosity (dl/g)	M _w (approx.)
12	23.0	18.5	50/50	920 µl	0.126*	low
13	23.0	18.5	50/50	1380 µl	0.108*	low

M_w are relative to polystyrene standard

* are reduced specific viscosities of a 1 g/100 ml solution in chloroform.

[0046] Alternatively, the lactide, glycolide and lactic acid if present, may be heated at 160°C and 0.08 g of stannous octanoate then added to initiate the polymerisation.

Example 14 (outside scope of invention)

[0047] A polylactide comprising equimolar proportions of glycolic acid and D,L-lactic acid units, and having an intrinsic viscosity of 1.36 (50 mg) was dissolved in dioxan (1 ml), and 50 µl of a solution of ICI.118630,



(233 mg per ml of the acetate salt, equivalent to 200 mg per ml of base) in distilled water was added. The resultant hazy solution was cast as a film, the solvents were evaporated in a stream of nitrogen in the dark, and the film was dried at 40°C under reduced pressure [0.027 mbar (0.02 mm of mercury)] for 48 hours. The mixture, containing ~17% by weight of ICI.118630 in the polylactide was homogenized by three successive compression mouldings at 110°C for 10 seconds, and was finally compression moulded into implants 0.038 cm thick, each weighing 1.5 mg and containing 309±7 µg (~17% by weight) of ICI.118630.

[0048] The continuous release of ICI.118630 from such implants was demonstrated by placing them in female rats exhibiting normal oestrous behaviour. After implantation, the rats went into a period of dioestrus, detected by the lack of cornified vaginal smears, lasting from 31 to 40 days, thus indicating that ICI.118630 was being continuously released during that period.

[0049] The process described above was repeated, using 50 µl of ICI.118630 acetate solution (150 mg of pure peptide per ml of water) and implants were made similarly, weighing 2 mg, containing 306±6 µg of ICI.118630 (13% by weight), and 0.038 cm in thickness. In the rat oestrus test described above, these implants released ICI.118630 continuously over a period of 30 to 38 days, as evidenced by the period of dioestrus in the rats.

[0050] For use in human therapy, implants containing 1 to 100 mg of ICI 118630 (5—50% by weight), weighing 2 mg—1 g, and in the form of cylindrical rods suitable for implantation by trochar, were manufactured by the process described above.

Example 15

[0051] The process described in Example 14 was repeated, but using polylactides comprising equimolar proportions of D,L-lactic acid and glycolic acid units, and having intrinsic viscosities of 0.33 and 0.25, instead of 1.36, to prepare implants containing 10% by weight of ICI.118630, weighing about 3 mg, and 0.08 cm thick.

[0052] These implants were placed in female rats (5 per group) which, prior to implantation, exhibited regular oestrous behaviour. The implants prepared from polylactide of intrinsic viscosity 0.33 exhibited an induction period of 5 days followed by a dioestrus period of about 26 days; and the implants prepared from polylactide of intrinsic viscosity 0.25 exhibited an induction period of 3—4 days, followed by a dioestrus period of about 25 days.

[0053] Similar implants, but containing 20% by weight of ICI.118630 were prepared in the same way, and these exhibited a similar dioestrus period, but no induction period, in the rat test described above.

[0054] For use in human therapy, implants containing 1 to 100 mg of ICI.118630 (5—50% by weight), weighing 2 mg—1 g, and in the form of cylindrical rods suitable for implantation by trochar were manufactured similarly.

Example 16 (outside scope of invention)

[0055] The process described in Example 14 was repeated, using a polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units, and having intrinsic viscosity of 1.36, to prepare mixtures of ICI.118630 and

polylactide containing 3% and 1% by weight of ICI.118630. The mixture was micronised at room temperature in an ultracentrifuge mill fitted with a 125 μ m (120-mesh) screen, and the micronised particles were suspended in propylene glycol for injection at a concentration of 100 mg per ml.

[0056] Female rats showing regular oestrous behaviour were injected subcutaneously with 0.1 ml of the 3% by weight propylene glycol suspension described above, or with 0.3 ml of the 1% suspension. Both groups were monitored, by examining vaginal smears daily for cornification, and exhibited occasional cornified smears up to 20 to 24 days after dosing, followed by a clear dioestrus period up to days 38 to 42 after dosing, thus demonstrating continuous release of ICI.118630 over that period.

Example 17 (outside scope of invention)

[0057] Tetragastrin hydrochloride, (Trp-Met-Asp-Phe-NH₂ · HCl), (200 mg) was dissolved in a mixture of dioxan (9 ml) and water (1 ml), and to the solution was added a polylactide as described in Example 11 (1.8 g). The mixture was cast as a film, and the solvents were evaporated in a stream of nitrogen. The resulting film was dried under reduced pressure [0.027 mbar (0.02 mm of mercury)] at 40°C for 48 hours, then homogenised by three successive compression mouldings at 80°C for 10 seconds, and moulded to give films of thickness 0.02, 0.06 or 0.12 cm, each weighing about 80 mg.

[0058] The release of tetragastrin was measured by placing a film in distilled water, removing a sample of the distilled water daily, replacing all the remaining distilled water by fresh, and measuring the ultraviolet absorption of the daily samples at 277 nm. The following results were obtained, which demonstrated continuous release of tetragastrin from films of all three thicknesses:—

Time (days)	Cumulative % tetragastrin released		
	0.02 cm film	0.06 cm film	0.12 cm film
1	9.6	5.6	4.0
4	14.9	10.5	9.0
7	20.3	13.9	11.7
9	25.3	17.7	14.8
11	33.1	22.6	19.4
14	48.6	33.2	28.1
17	61.9	45.9	40.5
21	74.7	59.8	53.2
24	81.8	68.1	60.2
28	85.2	74.9	66.4
31	86.9	77.7	70.9
36	88.5	82.5	77.5
39	89.2	85.1	82.6

Example 18 (outside scope of invention)

[0059] Tetragastrin hydrochloride (40 mg) was dissolved in aqueous dioxan (1:9 by volume), and added to a solution of:

(a) a polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units, and having a reduced specific viscosity of 0.115 (as a 1 g/100 ml solution in chloroform), (120 mg), and

(b) a polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid residues, and having an intrinsic viscosity of 1.08 (40 mg), in dioxan (2 ml). The mixed solutions were cast as a film, as described in Example 14, and moulded as implants weighing about 50 mg, and 0.02 cm thick.

[0060] The release of tetragastrin from these implants was measured by the procedure described in Example 17, and the following results showing continuous release were obtained:—

Time (days)	Cumulative % tetragastrin released
1	0.6

(continued)

Time (days)	Cumulative % tetragastrin released
2	1.0
3	1.6
4	2.7
7	8.6
10	17.2
14	29.4
18	43.1
23	56.3
28	64.9
32	71.2
36	79.0
39	83.9
44	90.5

Example 19 (outside scope of invention)

[0061] A polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units, and having an inherent viscosity of 0.11 as a 1 g/100 ml solution in chloroform, (50 mg), was dissolved in dioxan (1 ml) and a solution of mouse epidermal growth factor (EGF, 0.05 mg) in water 0.05 ml) was added. The mixture was cast as a film on polytetrafluoroethylene cloth, and the solvent was removed in a stream of nitrogen in the dark. The film was dried at 60°C, under reduced pressure [0.11 mbar (0.8 mm of mercury)] for 48 hours. The film was then compression moulded at 120°C for 10 seconds to give implants 0.02 cm thick, weighing 10 mg.

[0062] The implants were each placed in a black vial at 37°C with 1 ml of McIlvain's buffer (pH 7.4), (Documenta Geigy, Scientific Tables, edited by K. Diem and C. Leutner, published by J. R. Geigy SA, Basle, Switzerland, 7th Edition, 1970, page 280), containing 0.02% by weight of sodium azide. The buffer was removed daily, and replaced with fresh, and the EGF released into the buffer by the implant was measured by radio-immuno assay, which demonstrated that release started immediately, and continued for at least 2 weeks releasing 100—200 µg per day.

Example 20 (outside scope of invention)

[0063] A polylactide comprising equimolar proportions of D,L-lactide and glycolide and having a reduced specific viscosity (1 g/100 ml solution in chloroform) of 0.11, (400 mg), was dissolved in dioxan (2 ml), and a solution/suspension of bovine prolactin (100 mg) in distilled water (0.5 ml) was added, with vigorous agitation. The mixture was poured onto a polytetrafluoroethylene cloth, and dried, first in a stream of nitrogen and then under reduced pressure [0.013 mbar (0.01 mm of mercury)] at 40°C for 24 hours. The heterogeneous mixture thus obtained was homogenised by four successive compression mouldings at 60°C, and then moulded into a slab 0.2 cm thick, from which implants weighing 60 mg were excised.

[0064] The implants were placed subcutaneously into adult female rats, which were then periodically bled from the tail, and the prolactin in the blood samples thus obtained was measured by radio-immune assay. A placebo group, receiving no prolactin in the implant, was similarly assayed, and the levels of circulating prolactin compared. The following results were obtained:—

Time	Plasma level of bovine prolactin (µg/ml)	
	Placebo group	Treatment group
1	0.38	24.7
2	0.45	105.9
6	0.54	7.7
9	0.72	17.8
13	0.52	65.4
16	0.56	89.7
20	0.75	288

(continued)

Time	Plasma level of bovine prolactin ($\mu\text{g/ml}$)	
	Placebo group	Treatment group
23	0.81	142
26	0.84	562
42	1.25	1250

Example 21 (certain examples outside scope of invention as specified in Examples 1 to 13)

[0065] The process described in Examples 1—13 was repeated, except that the polylactide was dissolved in dioxan instead of in chloroform, and similar polylactides were obtained.

Examples 22—29

[0066] The process described in Examples 1—13 was repeated, except that the polylactide was dissolved in glacial acetic acid, and the glacial acetic acid solution thus obtained was added dropwise to methanol in order to precipitate the polylactide, which was filtered off and dried under vacuum at 40°C for 24 hours, then at 80°C for 24 hours.

[0067] The following particular polylactides were prepared by this method:—

Ex.	D,L-lactide (L) (g)	Glycolide (G) (g)	L/G Molar proportion	D,L-lactic acid	Intrinsic viscosity	Mw. (approx.)	Stannous octanoate
22	33.3	26.7	50/50	360 µl	0.234	13,500	93 µl
23	11.1	8.9	50/50	120 µl	0.243	14,450	31 µl
24	11.1	8.9	50/50	120 µl	0.265	16,800	31 µl
25	111.0	89.0	50/50	1.2 ml	0.257	15,850	0.31 ml
26	111.0	89.0	50/50	1.2 ml	0.239	14,200	0.31 ml
27	88.8	71.2	50/50	0.86 ml	0.262	16,600	0.25 ml
28	22.2	17.8	50/50	1.38 ml	0.09*		62 µl
29**	20.0	—	100/0	0.12 ml	0.260	16,200	31 µl

* Inherent viscosity as 1 g/100 ml solution in chloroform.

** Outside scope of invention

Example 30

[0068] A polylactide comprising equimolar proportions of glycolic acid and D,L-lactic acid units and having an intrinsic viscosity of 0.25 was dissolved in glacial acetic acid, and the solution was freeze-dried. The freeze dried powder (540.7 mg) and ICI 118630 (142.1 mg) of the acetate salt (equivalent to >124 mg of base) were dissolved in 6.8 ml of acetic anhydride-free glacial acetic acid and freeze dried for 24 h.

[0069] (The glacial acetic acid was refluxed for 2 h with 1% water to remove the acetic anhydride): The freeze dried product was extruded under pressure at 70°C to a 1 mm diameter rod, from which implants of the required weight were cut. The implants were dissolved in an appropriate solvent, for example acetonitrile, and assayed for drug content and purity. Implants were shown to contain 16.1% w/w pure 118630 base.

[0070] Release of 118630 was evaluated by immersing implants weighing approximately 10 mg in McIlvains pH 7.4 buffer at 37°C. ICI 118630 was released continuously for at least 5 weeks.

[0071] In a further experiment, implants weighing approximately 390 µg, 860 µg, 1500 µg, 3000 µg and ~6000 µg

were implanted subcutaneously in groups of adult, regularly cycling female rats. In the 28 days following implantation, animals were essentially free of oestrus intervals showing that active drug was released continuously over this period.

Example 31

[0072] A solution of ICI 118630 acetate salt was prepared by dissolving 170.8 mg of ICI 118630 acetate in 5 ml of acetic anhydride-free glacial acetic acid. (The glacial acetic acid was fluxed for 2 h with 1% water to remove acetic anhydride). This solution was shown by high pressure liquid chromatography (HPLC), to contain 25.21 mg of ICI 118630 base per ml, 442.5 mg of polylactide (prepared as in Example 25 was dissolved in 4.5 ml of the acetic acid solution, and the resulting solution was freeze dried for 25 h. The freeze dried product was extruded under pressure at 74°C to a 1 mm diameter rod, from which implants of the required weight were cut. The implants were dissolved in an appropriate solvent, such as acetonitrile, and the resulting solution was analysed by HPLC. The implants were shown to contain 20% w/w pure 118630 base.

Example 32 (outside scope of invention)

[0073] A polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units and having a reduced specific viscosity of 0.126 (1 g/100 ml solution in chloroform), (240 mg), was dissolved in glacial acetic acid (5 ml) and a solution of tetragastrin hydrochloride (60 mg) dissolved in glacial acetic acid (5 ml) was added with vigorous agitation. The solution was freeze dried for 24 h and the resultant solid was compression moulded at 50°C for 20 seconds to give implants 0.2 cm thick and weighing 35—40 mgs.

[0074] The above procedure was repeated with the following polymers:—

(a) a polylactide comprising 67 mole % of D,L-lactic acid and 33 mole% glycolic acid units and having a reduced specific viscosity of 0.121 (1 g/100 ml solution in chloroform).

(b) a polylactide comprising 75 mole % of D,L-lactic acid and 25 mole % glycolic acid units and having a reduced specific viscosity of 0.108 (1 g/100 ml solution in chloroform).

(c) a polylactide comprising 100% of D,L-lactic acid and having a reduced specific viscosity of 0.100 (1 g/100 ml solution of chloroform).

[0075] The release of tetragastrin from these implants was measured by the procedure described in Example 17, and the following results were obtained:—

Time (days)	Cumulative % tetragastrin released			
	50% D,L-lactide 50% glycolide	67% D,L-lactide 33% glycolide	75% D,L-lactide 25% glycolide	100% D,L-lactide
1	6.0	1.0	1.7	1.9
3	12.9	2.1	3.2	3.0
7	23.3	7.1	8.0	5.1
9	27.2	12.2	11.6	6.5
11	30.3	20.2	15.8	8.2
15	36.7	40.0	27.0	14.0
17	39.3	45.0	29.7	17.9
21	44.5	51.8	35.1	25.5
24	49.6	55.6	37.9	30.4
28	58.8	59.6	41.1	36.1
31	68.8	62.3	42.8	40.1
35	81.5	67.3	45.0	45.6
39	91.0	74.3	47.6	50.7
42	95.9	81.9	50.6	55.3
46	96.5	89.1	55.5	60.4
49	97.5	93.2	60.0	65.2
53		95.4	64.8	70.0
56		96.3	68.6	73.6

(continued)

Time (days)	Cumulative % tetragastrin released			
	50% D,L-lactide 50% glycolide	67% D,L-lactide 33% glycolide	75% D,L-lactide 25% glycolide	100% D,L-lactide
59		97.0	73.1	77.8
63		97.2	77.1	81.5
70			82.7	86.1
74			85.0	87.5
84			90.4	89.5

[0076] These results show that continuous release of tetragastrin is obtained using low molecular weight polylactides and that duration of release is determined by the composition of the hydrolytically unstable polyester.

Example 33 (outside scope of invention)

[0077] A polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units and having an inherent viscosity of 0.126 (as a 1 g/100 ml solution in chloroform) (9.5 mg), was dissolved in distilled dioxan (0.25 ml) and a solution of mouse epidermal growth factor (EGF 0.5 mg) in distilled water (0.1 ml) was added. The mixture was cast as a film on polytetrafluoroethylene cloth, and the solvent was removed in a stream of nitrogen, in the dark. The film was dried at 40°C under reduced pressure [0.013 mbar (0.01 mm of mercury)] for 48 hrs. The film was then compression moulded at 70°C for 10 seconds to an implant 0.02 cm thick, weighing about 9 mg. A placebo implant was also prepared.

[0078] The samples were implanted subcutaneously into carotid cannulated guinea pigs, blood samples taken periodically and the plasma EGF levels were measured by radio-immuno assay.

[0079] Raised plasma EGF levels were observed from Day 3 and continued for at least 1 week.

[0080] Similar implants were prepared as described above, but using a polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units and having an intrinsic viscosity of 1.06. Compression moulding of this implant was done at 120°C.

[0081] Implantation and plasma assays were done as described above, but raised plasma EGF levels were not observed until day 17 after implantation.

Example 34 (outside scope of invention)

[0082] A polylactide comprising equimolar proportions of D,L-lactic acid and glycolic acid units and having an inherent viscosity of 0.093 as a 1 g/100 ml solution in chloroform (40 mg) was dissolved in anhydrous-free glacial acetic acid (1 ml) and a solution of mouse epidermal growth factor (EGF, 8.15 mg) in a mixture of water (0.5 ml) and anhydride-free glacial acetic acid (3 ml) was added. The solution was freeze-dried for 24 h. The resulting powder was then compression moulded at 50°C to give an implant 2 mmx2 mmx 10 mm weighing 36.1 mg.

[0083] The sample was implanted subcutaneously into a cannulated cat, blood samples were taken periodically and the plasma EGF levels were measured by radio-immuno assay.

[0084] Raised plasma EGF levels were observed from day 3 and continued for at least 40 days.

Example 35 (outside scope of invention)

[0085] Implants containing bovine prolactin were prepared as described in Example 20, but using:—

- (a) a polylactide (400 mg) comprising equimolar proportions of D,L-lactic acid and glycolic acid units and having a reduced specific viscosity (1 g/100 ml solution in chloroform) of 0.11, dissolved in 4 ml dioxan; and
- (b) a polylactide (400 mg) comprising equimolar proportions of D,L-lactic acid and glycolic acid units and having an intrinsic viscosity of 1.06 dissolved in 4 ml dioxan. This sample was moulded at 110°C.

[0086] Formulations (a) and (b) were each tested *in vivo* as described in Example 20. Formulation (a) released significant levels of plasma bovine prolactin from at least as early as day 4 and continued to release for at least 85 days, while formulation (b) released significant levels from at least as early as day 8 for at least 85 days.

Claims

1. A solid pharmaceutical composition for subdermal implantation comprising a polylactide, which is a copolymer of lactic and glycolic acids made by a ring opening, polymerisation of a mixture of cyclic dimer of lactic acid and cyclic dimer of glycolic acid in the presence of chain stopping agent or a mixture of such copolymers, and an acid stable polypeptide which is not significantly hydrolysed under the conditions encountered within the composition during the period of use envisaged, which composition, when placed in an aqueous physiological-type environment, exhibits a release profile which has two successive phases of release of the polypeptide as an aqueous solution, the first phase being released by matrix diffusion and the second phase being released consequent upon degradation of the polylactide until essentially all of the polypeptide has been released, **characterised in that** the diffusion phase and the degradation phase of the release profile overlap in time, and the release of the polypeptide occurs over a period of at least one week, the composition being adapted to achieve the release profile by varying the polylactide composition, particularly the proportion of lactic acid to glycolic acid, by choosing the weight average molecular weight of the polylactide and its polydispersity, by choosing the proportion of the polypeptide to polylactide or by choosing the geometry of the solid formulation for implantation to provide the release profile when taking account of the molecular weight of the polypeptide and interaction of basic polypeptides with the terminal carboxylic-acid groups of the polylactide and wherein either (a) the composition comprises from 5 to 50% by weight of ICI 118,630

(Pyro-Glu-His-Trp-Ser-Tyr-D-Ser (O-tBu) -Leu-Arg-Pro-Azgly-NH₂)

and from 50 to 95% by weight of polylactide wherein the ratio of glycolide to lactide units is from 0.2 to 3, and which has an inherent viscosity of 0.2 to 0.5 dl/g (1g per 100ml in chloroform) or (b) the composition comprises from 0.1 to 50% by weight of ICI 118,630

(Pyro-Glu-His-Trp-Ser-Tyr-D-Ser (O-tBu) -Leu-Arg-Pro-Azgly-NH₂)

and from 50 to 99.9% by weight of a polylactide wherein the ratio of glycolide to lactide units is up to 3, and which has an inherent viscosity of less than 0.2 dl/g (1g per 100ml in chloroform) and excluding a composition in microcapsule form comprising at least one polypeptide which is a naturally occurring luteinising hormone releasing hormone (LH—RH), a synthetically prepared material of the same type or synthetically prepared analogues of naturally occurring LH—RH which act in some manner on the anterior pituitary gland to affect the release of luteinising hormone (LH) and follicle stimulating hormone (FSH).

Patentansprüche

1. Feste pharmazeutische Zusammensetzung für die subdermale Implantation, die ein Polylactid, das ein Copolymer von Milch- und Glykolsäure, das durch ringöffnende Polymerisation einer Mischung des cyclischen Dimeren von Milchsäure und des cyclischen Dimeren von Glykolsäure in Gegenwart eines Kettenabbruchmittels hergestellt wurde, oder eine Mischung derartiger Copolymerer ist, und ein säurestabiles Polypeptid, das unter den Bedingungen, die innerhalb der Zusammensetzung während des beabsichtigten Verwendungszeitraums angetroffen werden, nicht nennenswert hydrolysiert wird, umfaßt, wobei die Zusammensetzung, wenn sie in einer wäßrigen Umgebung vom physiologischen Typ angeordnet wird, ein Freisetzungsprofil zeigt, das zwei aufeinanderfolgende Freisetzungsphasen für das Polypeptid in Form einer wäßrigen Lösung aufweist, wobei die erste Phase durch Matrixdiffusion freigesetzt wird und die zweite Phase anschließend durch Abbau des Polylactids freigesetzt wird, bis im wesentlichen das gesamte Polypeptid freigesetzt ist, **dadurch gekennzeichnet, dass** die Diffusionsphase und die Abbauphase des Freisetzungsprofils zeitlich überlappen, und die Freisetzung des Polypeptids über einen Zeitraum von wenigstens einer Woche erfolgt, wobei die Zusammensetzung dafür ausgelegt ist, dieses Freisetzungsprofil durch Variation der Polylactidzusammensetzung zu erreichen, und zwar insbesondere des Verhältnisses von Milchsäure zu Glykolsäure, indem man das gewichtsmittlere Molekulargewicht des Polylactids und dessen Polydispersität wählt, das Verhältnis des Polypeptids zu dem Polylactid wählt oder die Geometrie der festen Formulierung für die Implantation wählt, um das Freisetzungsprofil zu erhalten, wenn man das Molekulargewicht des Polypeptids und die Wechselwirkung von basischen Polypeptiden mit den terminalen Carbonsäuregruppen des Polypeptids berücksichtigt, und wobei entweder (a) die Zusammensetzung von 5 bis 50 Gew.-% von ICI 118,630

(Pyro-Glu-His-Trp-Ser-Tyr-D-Ser (O-tBu) -Leu-Arg-Pro-Azgly-NH₂)

und von 50 bis 95 Gew.-% Polylactid umfaßt, wobei das Verhältnis von Glycolid- zu Lactid-Einheiten von 0,2 bis 3 beträgt, und eine inhärente Viskosität von 0,2 bis 0,5 dl/g (1 g pro 100 ml in Chloroform) aufweist, oder (b) die Zusammensetzung von 0,1 bis 50 Gew.-% ICI 118,630

(Pyro-Glu-His-Trp-Ser-Tyr-D-Ser (O-tBu) -Leu-Arg-Pro-Azgly-NH₂)

und von 50 bis 99,9 Gew.-% eines Polylactids umfaßt, wobei das Verhältnis von Glycolid- zu Lactid-Einheiten bis zu 3 beträgt, und eine inhärente Viskosität von weniger als 0,2 dl/g (1 g pro 100 ml in Chloroform) aufweist, und wobei eine Zusammensetzung in Mikrokapselform ausgeschlossen ist, die wenigstens ein Polypeptid umfaßt, das ein natürlich vorkommendes Release-Hormon eines luteinisierenden Hormons (LH-RH), ein synthetisch hergestelltes Analoges des natürlich vorkommenden LH-RH ist, die auf irgendeine Weise so auf den Hypophysenvorderlappen wirken, dass die Freisetzung von luteinisierendem Hormon (LH) und follikelstimulierendem Hormon (FSH) bewirkt wird.

Revendications

1. Composition pharmaceutique solide pour implantation sous le derme, comprenant un polylactide, qui est un copolymère d'acide lactique et d'acide glycolique produit par polymérisation, par ouverture de cycle, d'un mélange d'un dimère cyclique d'acide lactique et d'un dimère cyclique d'acide glycolique en présence d'un agent de terminaison de chaîne, ou un mélange de tels copolymères, et un polypeptide stable en milieu acide, qui n'est pas hydrolysé de manière significative dans les conditions rencontrées dans la composition au cours de la période d'utilisation envisagée, composition qui, lorsqu'elle est placée dans un environnement aqueux de type physiologique, présente un profil de libération qui comporte deux phases successives de libération du poly-peptide sous forme d'une solution aqueuse, la première phase étant une phase de libération par diffusion à travers la matrice et la seconde phase étant une phase de libération suite à la dégradation du polylactide jusqu'à ce que pratiquement la totalité du polypeptide ait été libérée, **caractérisée en ce que** la phase de diffusion et la phase de dégradation du profil de libération se chevauchent dans le temps, et la libération du polypeptide se produit pendant une période de temps d'au moins une semaine, la composition étant adaptée à présenter le profil de libération en faisant varier la composition de polylactide, en particulier la proportion d'acide lactique par rapport à l'acide glycolique, en choisissant la moyenne en poids du poids moléculaire du polylactide et sa polydispersité, en choisissant la proportion du polypeptide par rapport au polylactide ou en choisissant la géométrie de la formulation solide pour implantation afin d'obtenir le profil de libération lorsqu'on prend en considération le poids moléculaire du polypeptide et l'interaction des polypeptides basiques avec les groupes acide carboxylique terminaux du polypeptide, et soit (a) la composition comprend 5 à 50 % en poids de ICI 118 630

(Pyro-Glu-His-Trp-Ser-Tyr-D-Ser (O-tBu) -Leu-Arg-Pro-Azgly-NH₂)

et 50 à 95 % en poids de polylactide dans lequel le rapport des motifs glycolide aux motifs lactide va de 0,2 à 3, et possède une viscosité inhérente de 0,2 à 0,5 dl/g (1 g pour 100 ml dans le chloroforme) soit (b) la composition comprend 0,1 à 50 % en poids de ICI 118 630

(Pyro-Glu-His-Trp-Ser-Tyr-D-Ser (O-tBu) -Leu-Arg-Pro-Azgly-NH₂)

et 50 à 99,9 % en poids de polylactide dans lequel le rapport des motifs glycolide aux motifs lactide va jusqu'à 3 et possède une viscosité inhérente inférieure à 0,2 dl/g (1 g pour 100 ml dans le chloroforme)

en excluant une composition sous forme de micro-capsules comprenant au moins un polypeptide qui est une hormone de libération d'hormone lutéinisante naturelle (LH-RH), une substance du même type préparée par synthèse ou des analogues préparés par synthèse de la LH-RH naturelle qui agissent d'une certaine manière sur

l'hypophyse antérieure en ayant un effet sur la libération d'hormone lutéinisante (LH) et la folliculostimuline (FSH).

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